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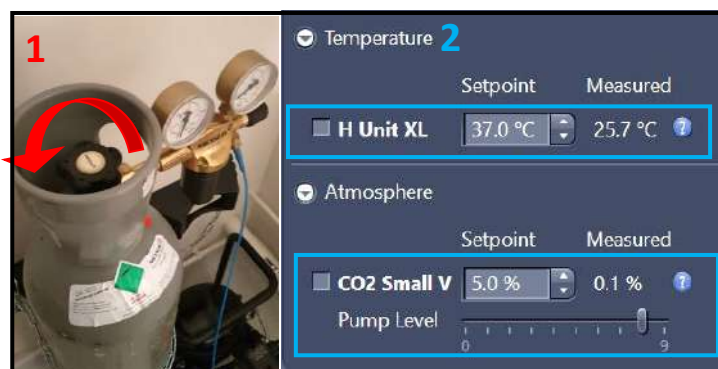
START THE SYSTEM

1. Press the « Components » button on the ON
2. Press the button on the ON to activate the LED used to the ocular observation.
3. Switch on the PC and login into the session « USER ».
4. Start the software by clicking on the ZEN blue The icon on the window desktop.
5. Click on the ZEN SYSTEM button to access the acquisition menu.



START THE TEMPERATURE AND CO2

1. Open the bottle of CO2
2. On the right side of the ZEN software, activate the temperature and CO2



OCCULAR OBSERVATION

1. Select the LOCATE tab (first on the left).
2. Select the desired illumination
3. Open/Close the transmission light
4. Open/close the reflected light
5. Adjust the fluorescent lamp intensity
6. Choose your objective
Depending on the type of illumination that you would like to.



Manual Command

1. Wheel to adjust the focus

PS: The big one permits you to adjust fast, and the extremity one permits you to adjust with more precision.

2. put down the objective to install your sample

3. Put back the objective at the focus point

4. Open/Close the shutter of the transmission light



ACQUIRE ONE OR SEVERAL COLORS

I. Dye Assistant

1. Select the **ACQUISITION** tab

2. Open SMART SETUP

3. Then, in the DYE menu select the different fluorophores and their respective colors

Several methods are suggested:

-**FASTEST:** Allows simultaneous acquisition for all your channels; this is the fastest method, but cross-talk between consecutive channels can be important.

-**BEST SIGNAL:** Each channel is acquired separately. This method is the slowest one but it avoids (as far as it is possible) cross-talk between consecutive channels.

-**SMARTTEST:** Compromise between speed and reduction of cross-talk. For example, both blue and red channels are acquired simultaneously, then the green one is acquired separately.

Remarks: These modes are just a base to start a configuration. The software doesn't necessarily choose the best one. You have to check by yourself and then modify appropriately the settings.

Dye	Wavelength (nm)	Color
5-TAMRA	567	Yellow
Alexa Fluor 633	647	Red
Cy3	561	Yellow
DAPI	465	Blue
EGFP	509	Green

II. Imaging Setup

1. In the IMAGING SETUP window, each TRACK corresponds to one sequence of one or several colors.

2. Add or remove sequences by clicking on +LSM Confocal/-

3. Adapt wavelength detection by adjusting the detection window for each channel.

4. Add or remove channels with +/- for each sequence.

5. Select, if possible, the same dichroic for each sequence to accelerate the change between sequences.

If no dichroic is required, select the « Plate » filter.

6. Activate the 405 dichroic for each sequence even if you don't need it.

7. Tick T-PMT to acquire transmission images in one of your sequences.

The screenshot shows the 'Imaging Setup' window. At the top, the title bar is labeled '1'. Below it, there are two track buttons: 'Track1 Confocal' and 'Track2 Confocal', with a '+LSM Confocal' dropdown menu labeled '2' next to them. A 'Switch track every' dropdown is set to 'Frame'. A spectral graph shows two emission spectra: a broad one from 400-600nm and a narrow one at 646-721nm. Below the graph are two detection window sliders labeled '3'. A 'Mirror' dropdown is set to 'Plate'. A table lists channels with checkboxes, dyes, colors, names, and ranges. The 'T-PMT' checkbox is highlighted with a pink box and labeled '7'. A '+' and '-' button is highlighted with a yellow box and labeled '4'. At the bottom, there are two dichroic filter/detector pairs: 'MBS 488/5... Visible Light' (labeled '5') and 'MBS -405 Invisible Lig...' (labeled '6'). A 'Reflection' checkbox is at the bottom right.

Use	Dye	Color	Name	Range
<input checked="" type="checkbox"/>	DAPI	Blue	DAPI	412-528nm
<input type="checkbox"/>		White	ChS1	
<input checked="" type="checkbox"/>	AF633	Red	AF633	646-721nm
<input type="checkbox"/>		White	ChA	
<input type="checkbox"/>		White	T-PMT	

III. Acquisition SetUp

1. In the “ACQUISITION MODE” window, choose the sampling of your image which has an impact on the resolution.

2. Zoom in If necessary, move your zoom zone to see your object of interest. If necessary, rotate your image

PS: Zoom can increase the photobleaching but can decrease the acquisition time

3. Choose the scan speed (the faster mode has the worst signal/noise ratio).

4. Bidirectional mode allows you to acquire 2 times faster.

PS: the bidirectional mod could create phase issues that need correction X in phase settings

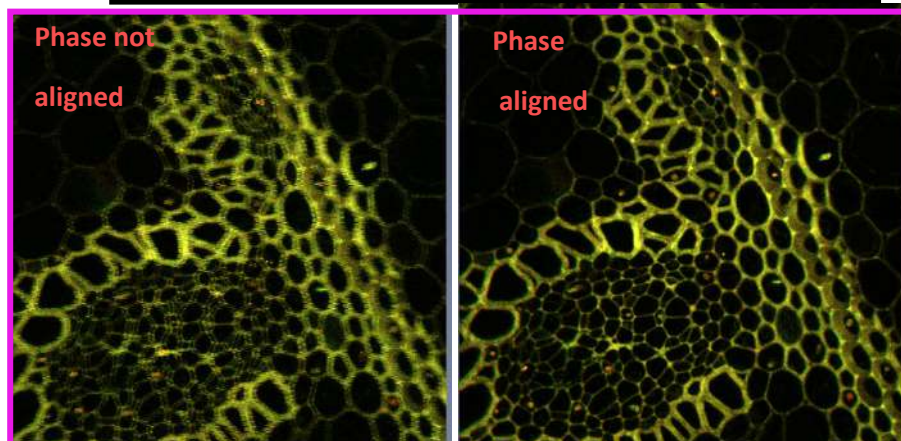
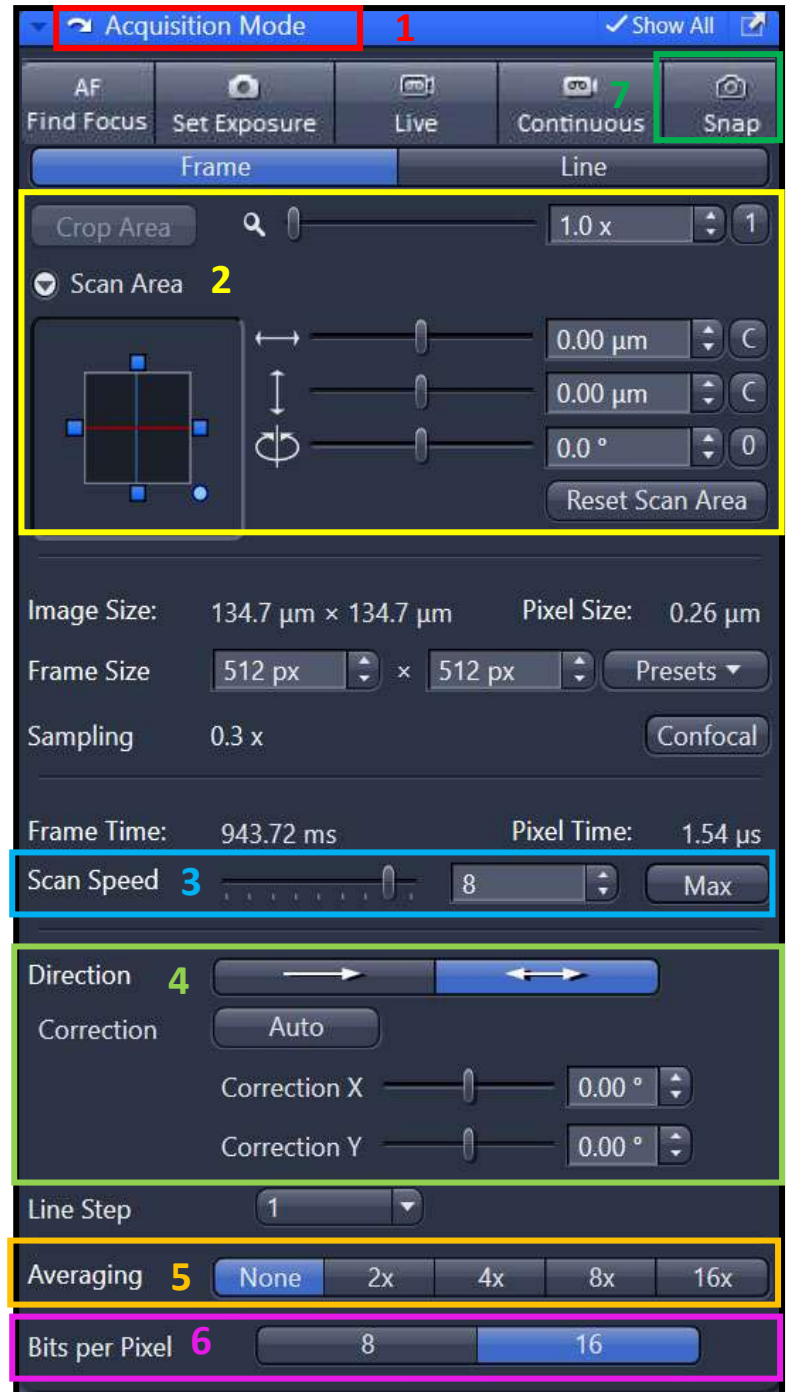
5. you can select the “AVERAGING” mode if the signal/noise ratio is not satisfying.

If you have a fixed sample the mode Frame is more appropriate. If you have a living sample the mode Line is preferable. (Method Mean is recommended)

6. Select 16 bits to increase your grey level and the quality of images.

7. Click on SNAP to acquire the image

PS: click on “Start Experiment” to begin your all acquisition (Tile; Z-stack etc).



IV. Channels

1. In the toolbox CHANNELS, adjust the pinhole at 1 AU (Airy unit) for each channel to obtain the best resolution/signal ratio.

2. Click on LIVE.

3. Adjust the laser power for each channel

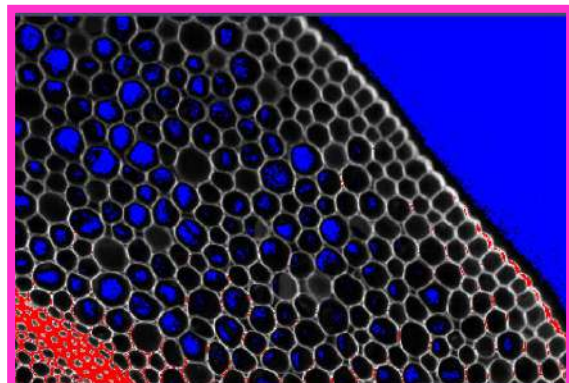
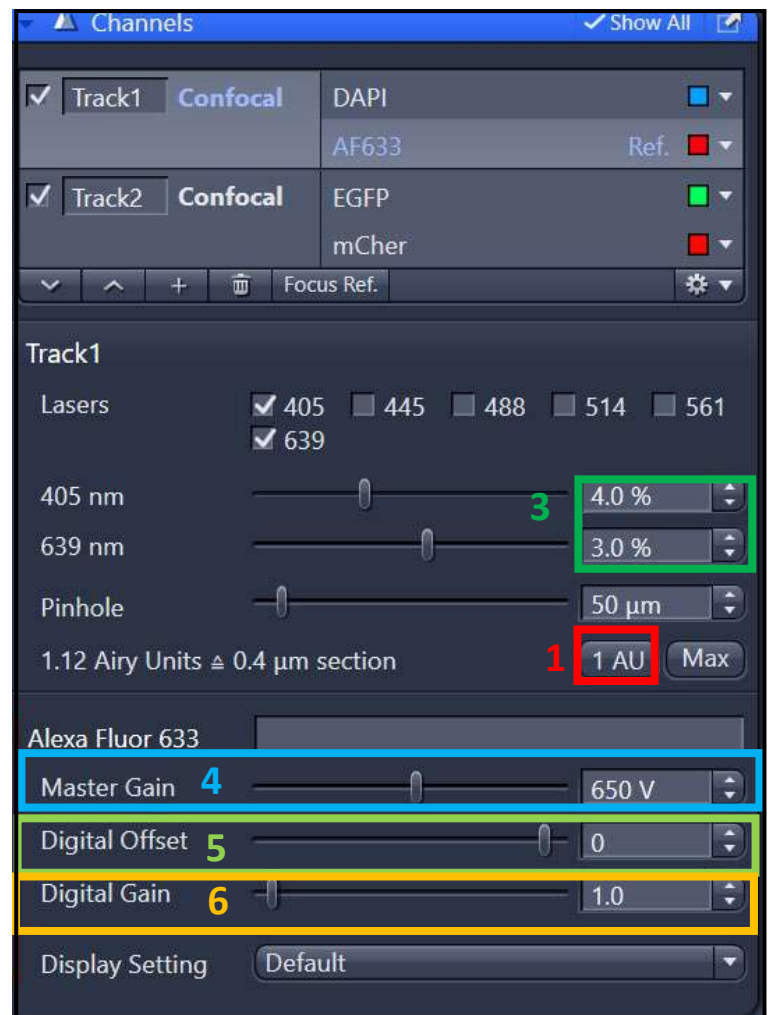
4. You can increase the GAIN (MASTER) to decrease the laser power if it is too high. But the signal/noise ratio will be impacted. **Stay between 650-800**

5. You can adjust the DIGITAL OFFSET to improve the image by removing the background signal

6. Don't increase the DIGITAL GAIN except if the GAIN (MASTER) is high enough

7. To find adapted parameters for gain and laser power, it is recommended to use the « Range Indicator » display.

In this representation, saturated pixels are red and black pixels are blue.



ACQUIRE IN AIRYSCAN 2

I. Dye Assistant

1. Select the ACQUISITION tab
2. Open SMART SETUP
3. Then in DYE select the different fluorochromes and their respective colors.
4. Select « Airyscan »

5. Several options are available:

Resolution: allows to acquire the optimal resolution of the Airyscan 2.

SNR/Sensitivity: Allows to acquire with the resolution of a confocal but with the sensitivity of the Airyscan

Speed: Allows to acquire using the fast modes (4Y or 8Y) of Airyscan 2.

Remarks: You can choose modes that make a compromise between the different options.

PS: The mode “Best or Smartest” is similar to the confocal mode.

These modes are just a base to start a configuration. The software doesn't necessarily choose the best one. You have to check by yourself and then modify appropriately the settings.

The screenshot displays the ZEN 3.1 (ZEN system) software interface. The top menu bar includes File, Edit, View, Acquisition, Graphics, Tools, Window, and Help. The toolbar below the menu bar contains icons for Locate, Acquisition (highlighted with a red box and labeled '1'), Processing, Analysis, and Applications. Below the toolbar, the 'Airyscan SR *' dropdown menu is open, showing 'Smart Setup' (labeled '2') and 'Reuse' buttons. The 'Add Dye or Contrasting Method' window (labeled '3') is open, showing a list of 'Recently Used' dyes with their respective wavelengths and colors: 5-TAMRA (567 nm, yellow), Alexa Fluor 633 (647 nm, red), Cy3 (561 nm, yellow), DAPI (465 nm, blue), EGFP (509 nm, green), and mCherry (610 nm, red). Below this list is a search bar. The 'Proposals' window (labeled '4') is open, showing a comparison between 'Best Signal' and 'Smartest (Live)' modes. The 'Smartest (Live)' mode is selected (labeled '5'). The 'Proposals' window displays two bar charts comparing 'Best Signal' and 'Smartest (Live)' modes across three channels (T1, T2, T3) for Contrast, SNR, and Speed. The 'Smartest (Live)' mode shows higher SNR and Speed compared to 'Best Signal'. Below the bar charts are three line graphs showing the signal profiles for each channel. The bottom of the window has checkboxes for 'Show Excitation' and 'Show Emission', and buttons for 'Reset', 'OK', and 'Cancel'.

II. Imaging Setup

1. Each “TRACK” represents a sequence of one or more colors.

2. Add or delete sequences by pressing +SR/-

3. Check/modify the filters selected for the acquisition of each Track.

PS: the filter selection corresponds to a combination of 2 filters.

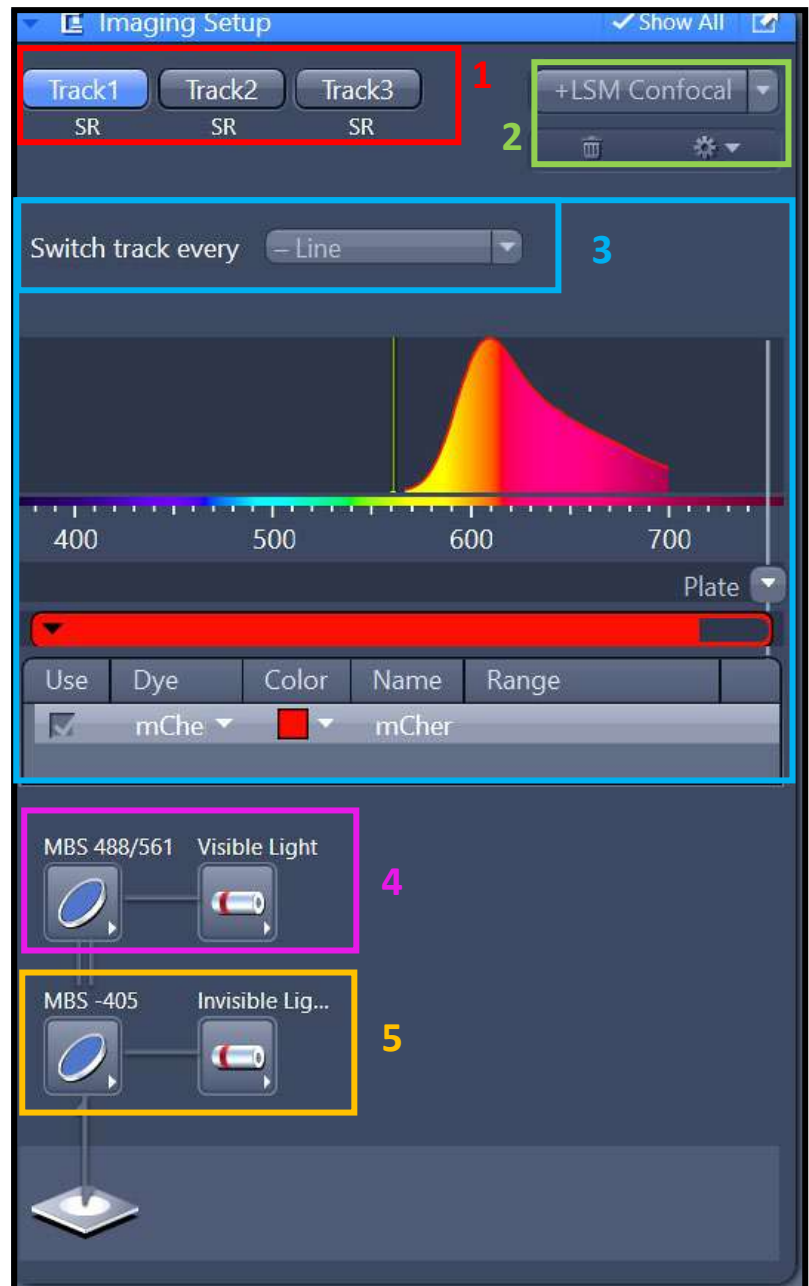
The spectrum acquires corresponding at the zone colored.

- If you use Line or Frame fast mode you have to choose the same filter combination for all the tracks.
- However, if you select the Frame mode you can set up a combination different for each filter.

4. Select if possible, the same dichroic for each sequence, to speed up the transition from one sequence to the other.

If you do not need to use a dichroic for an acquisition, you must choose the filter « plate ».

5. Activate the 405 dichroic for each sequence even if you do not use it.



II. ACQUISITION MODE

1. The Frame size defines your image resolution, but a high resolution involves a longer acquisition time.

PS: some Airy Scan mode has some definite parameters

- Select "**Confocal**" to keep the optimal resolution of a confocal.
- Select "SR" to maintain the optimal resolution of an airscan.

2. Modify the scan speed, a high speed decreased the signal/noise ratio (if you are "in vivo" select the Max)

3. Two-way mode allows you to go twice as fast.

PS: as in confocal mode the bidirectional mode could create phase issues. To resolve it you can correct it as explained previously (page 6, Number 4)

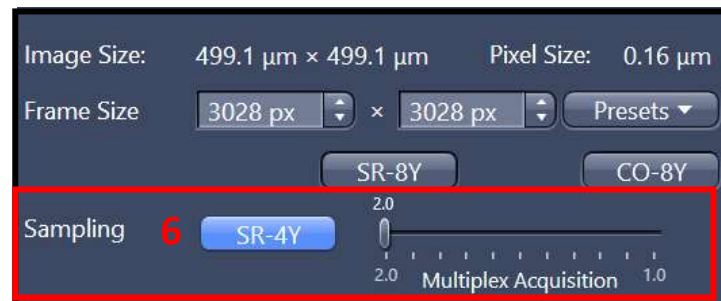
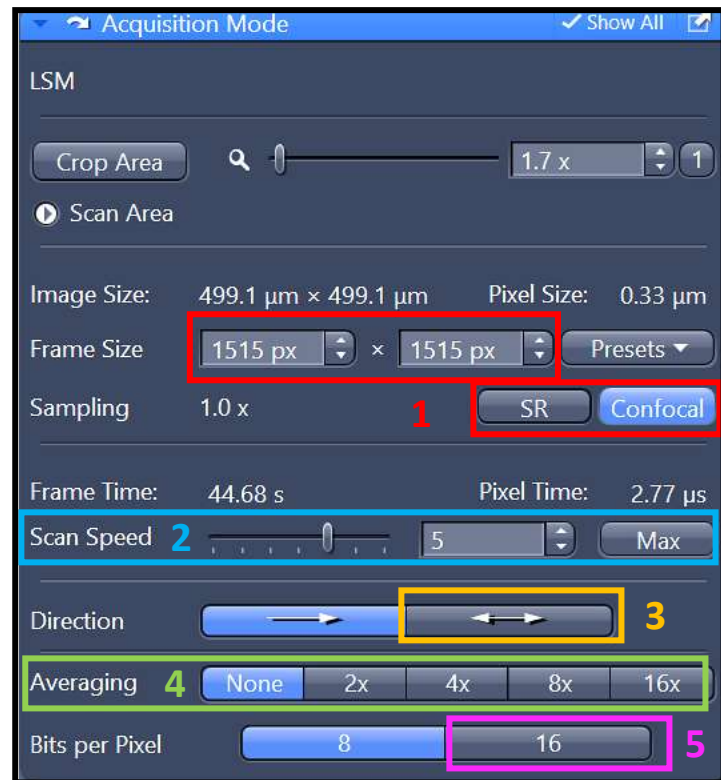
4. AVERAGING is a way to reduce noise ratio is not satisfactory. If the sample is fixed the Image mode is more appropriate. If the sample is alive the Line mode is preferable.

5. Select the 16 bits mode to obtain a better image quality.

6. in Fast SR, select the speed mode in the "Multiplex Acquisition" mode **SR-4Y**: SR reso with simultaneous 4-point scanning

7. Save your configuration.

PS: You can reuse the settings of an acquired image later by selecting the image and clicking on REUSE.



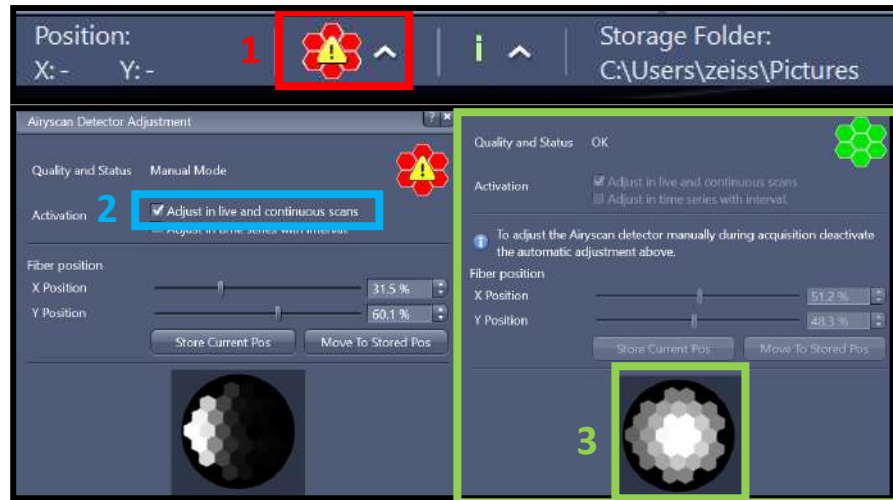
III. Set Up the Airy scan Sensor

1. To calibrate the AiryScan 2 sensors, start by selecting the red sensor icon.

2. On the new tab, select « Adjust in live and continuous scans ».

3. Select "Live" or "continuous scans". Wait until "quality and status" says "OK".

PS: Be sure that your signal intensity is high enough (you can increase the laser power or the gain).



4. Stop the acquisition, and uncheck the « Adjust in live and continuous scans » box.

IV. Channel Set Up

The channel setup is similar to the confocal mode explained on page 7

1. Adjust the laser power for each channel

The power of the laser is different in the confocal mode

2. In the toolbox CHANNELS, adjust the pinhole at 1 AU (Airy unit) for each channel to obtain the best resolution/signal ratio.

3. You can increase the GAIN (MASTER) to decrease the laser power if it is too high. But the signal/noise ratio will be impacted. Stay between 650-800

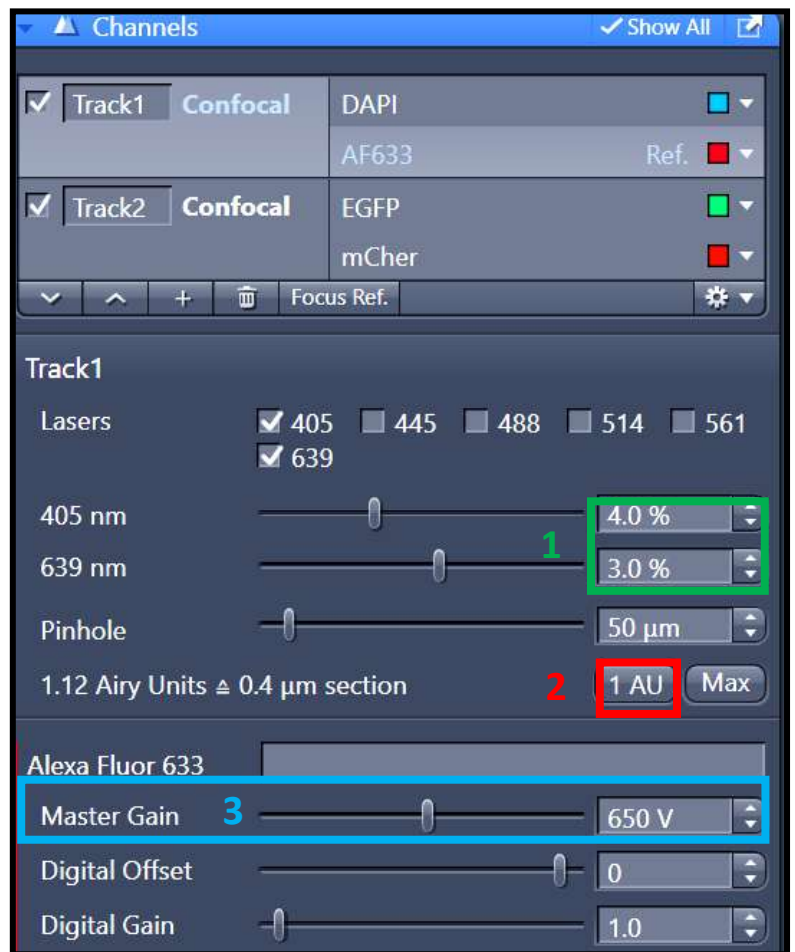


IMAGE PROCESSING.

1. Select the **PROCESSING** tab

2. In « *Method* » select « *Airyscan Processing* »

3. In « *Image Parameters* » select the image to be processed

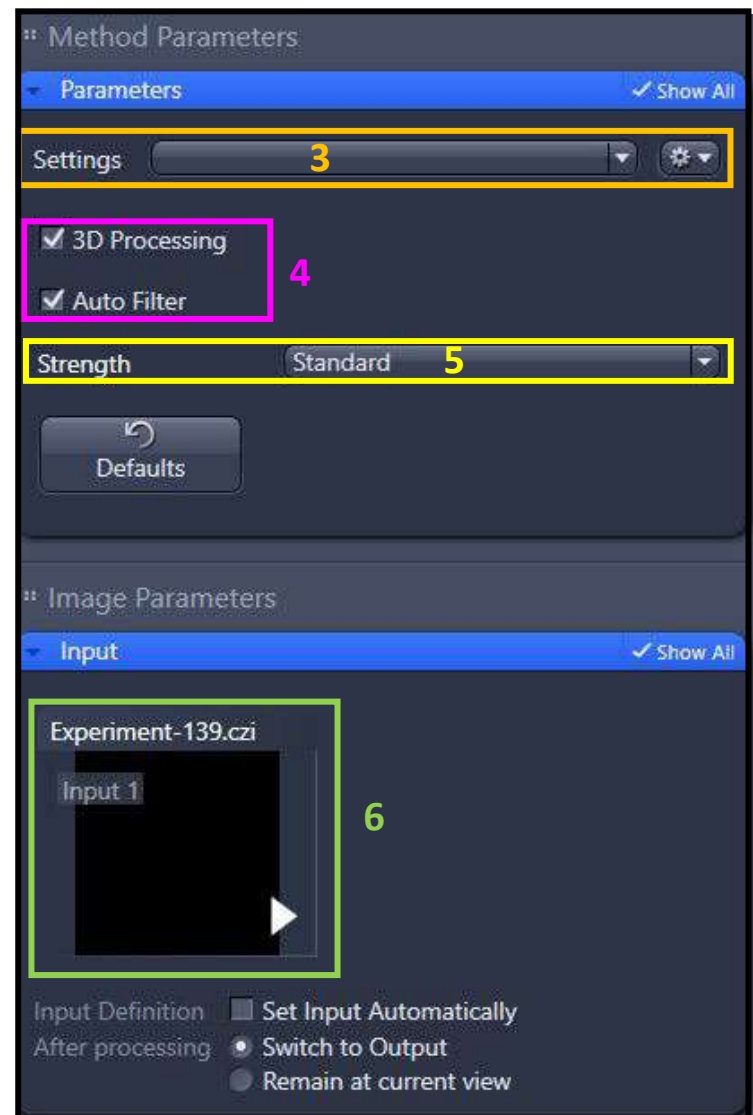
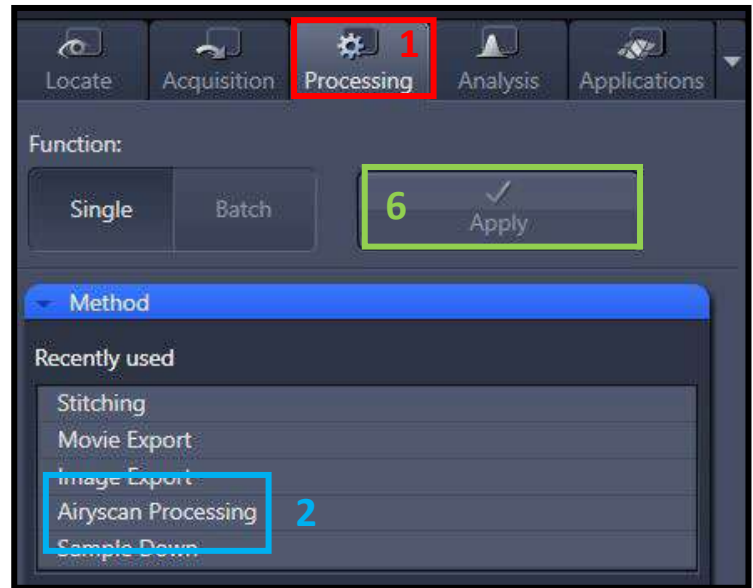
4. In « *Parameters* » tick « *Auto filter* »

When the acquisition is a Z-stack, the option is replaced by the option « *3D Processing* ». Tick this option when the Z interval is optimal.

5. Select the filtering mode from the 3 proposed:

- Low
- Standard
- Strong

6. In the « **Input** » Window choose the file that you want to process Click on « *Apply* »



ACQUIRE Z-STACK SERIES

1. Check the « Z-Stack » option.

2. Check « Show all Tools ».

3. For multicolor acquisitions, make the entire z-stack of one channel, then switch to the other one « Full Z-Stack per Channel » or all channels per plane « All Channels per Slices ».

Two acquisition mode exists: you can define the top and the bottom of your stack, or define only the center of your stack.

4. First /Last Mode

- Check the tab « *First/Last* »
- Make a « *Live* ».
- « *Set First* » defines one extremity of your stack and « *Set Last* » the other one

5. Define Center Mode

- Check the tab « *Center* ».
- Make a « *Live* ».
- Define the center of your stack with « *Center* ».

6. « *Interval* » has to be checked to ensure a fixed and chosen interval value in μm .

7. To choose the optimal resolution, you can use the « *Optimal* » option.

8. « *Range* » indicates the thickness of your stack.

9. The number of planes can be changed in « *Slices* ».

10. Start the acquisition by using « *Start Experiment* ».

The figure shows three screenshots of the software interface for acquiring a Z-stack series, with numbered callouts (1-10) highlighting specific settings and actions:

- Screenshot 1 (Top):** Shows the main acquisition control panel. Callout 1 points to the checked "Z-Stack" option. Callout 2 points to the "Show All" icon. Callout 3 points to the "Full Z-Stack per Channel" dropdown menu. Callout 4 points to the "Start Experiment" button.
- Screenshot 2 (Middle):** Shows the "Multidimensional Acquisition" window with the "Z-Stack" tab selected. Callout 4 points to the "First / Last" mode tab. Callout 5 points to the "Set Last" field (7546.82 μm). Callout 6 points to the "Interval" radio button. Callout 7 points to the "Optimal" resolution button (0.31 μm). Callout 8 points to the "Range" field (4.00 μm). Callout 9 points to the "Slices" field (5).
- Screenshot 3 (Bottom):** Shows the "Multidimensional Acquisition" window with the "Center" mode tab selected. Callout 5 points to the "Center" field (7544.82 μm). Callout 6 points to the "Slice" radio button. Callout 7 points to the "Optimal" resolution button (0.31 μm). Callout 8 points to the "Range" field (4.00 μm). Callout 9 points to the "Slices" field (5). Callout 10 points to the "Start Auto Configuration" button.

SPATIAL SAMPLING

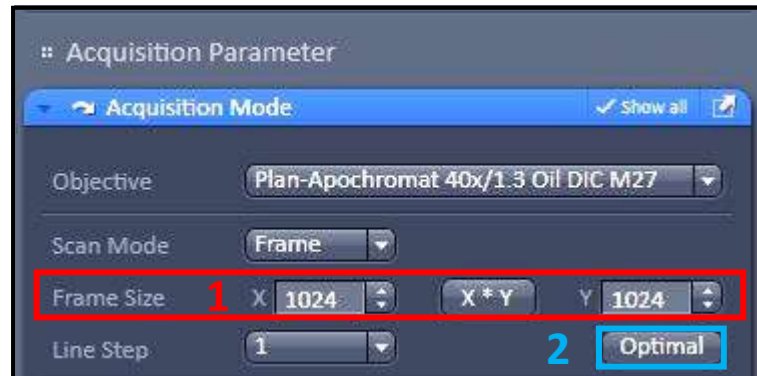
To obtain the optimal resolution for your image, the image voxel size must be equal to half of the resolution of the objective that you use (Nyquist criteria). On a laser scanning microscope, the lateral resolution (in XY) is better than the axial resolution (in Z).

I. Choose the pixel size by modifying

1. Sampling image means several pixels composing your image.
2. To choose automatically the best resolution depending on the objective and the zoom, click on OPTIMAL.

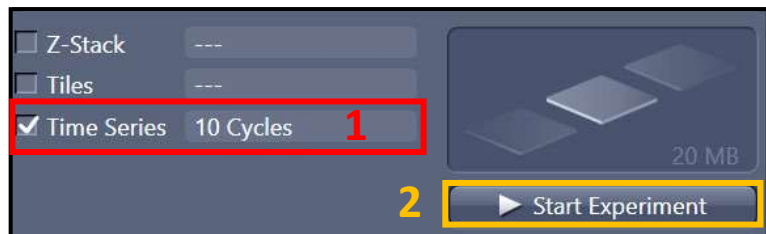
PS: Having pixel size smaller than the resolution is useless (over-sampling).

However, you can under-sample the image to increase the speed of the acquisition and decrease the loss of fluorescence because of the photobleaching effect.



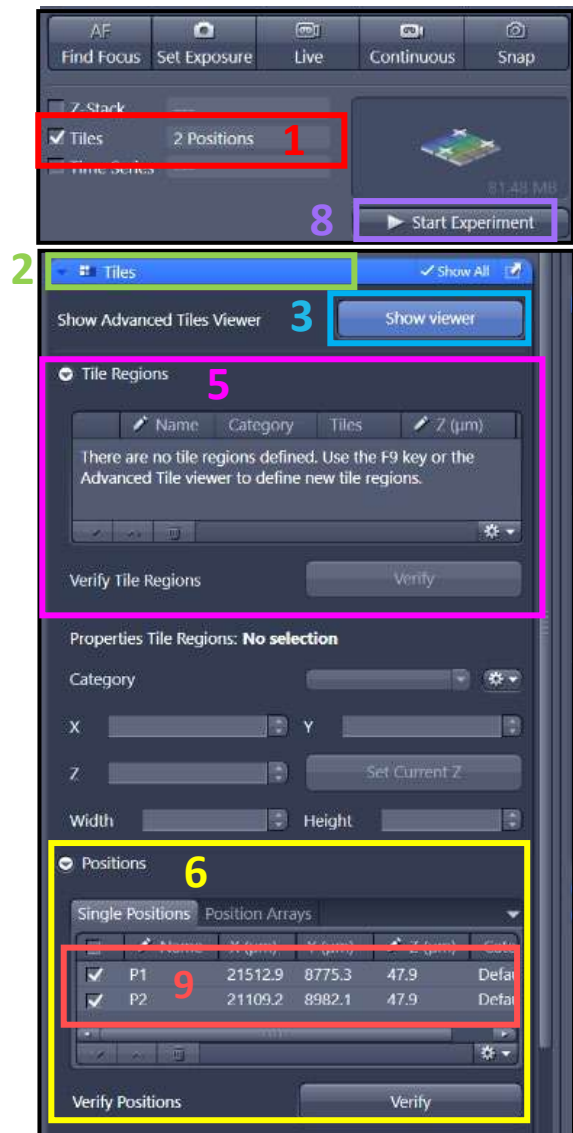
TEMPORAL SERIE

1. Select the **Time Series** box in the Acquisition Tab.
2. Choose the number of **cycles** and the **Interval** between each time point.
3. Click on **START EXPERIMENT** to start the acquisition



Tile scan/Multi-Position acquisition

1. Check « Tiles ».
2. Open the menu « Tiles ».
3. Click on « Show Viewer ». Make a « Live ».
4. The navigation space is displayed in the center of the software interface. Double-click where you want to acquire an image or use the joystick to move the stage
5. In the menu « Tile Regions », click on « Tiles » and double-click on your position. Your current position will correspond to the center of the tile.
6. Tile Position: you can add the position one by one to be more specific in your choice. For this, you have to Click on « + », of the position window to add the position selected on the navigation space
7. Click on « + or Add position/Tile», to validate the choice of your Region of Interest.
8. When all your positions and tile are selected you can click on « Start Experiment » to start the acquisition.

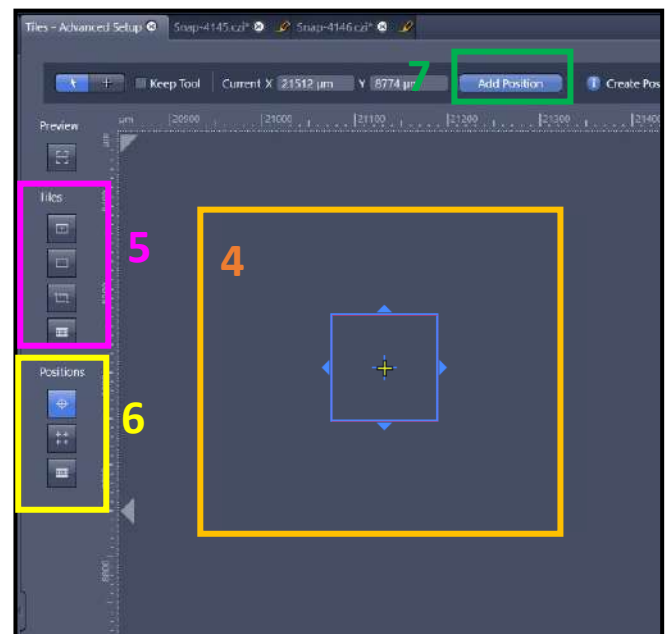


Focus correction

9. Make a right click on the position in the position window (tile window) and select "set current X/Y/Z or Current Z

PS: when you define the Z-stack it is important that you define with the Center mode as described previously to permit a z-stack adapted at the different Z set up of your different position.

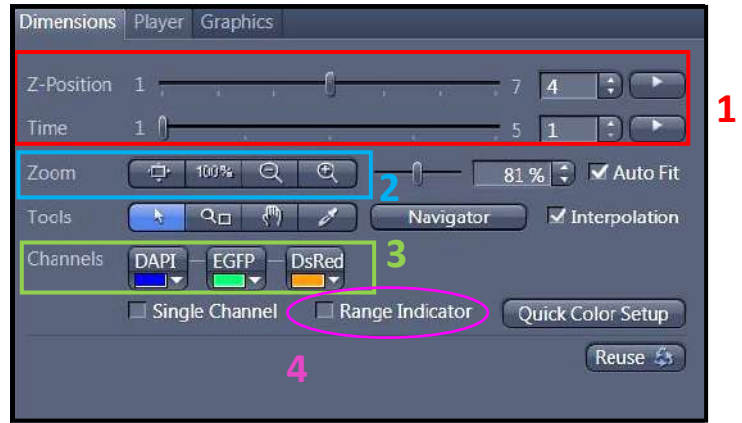
For specific operations, you can ask the engineers of the platform or ask for assistance hour on the microscope



GRAPHIC TOOLS

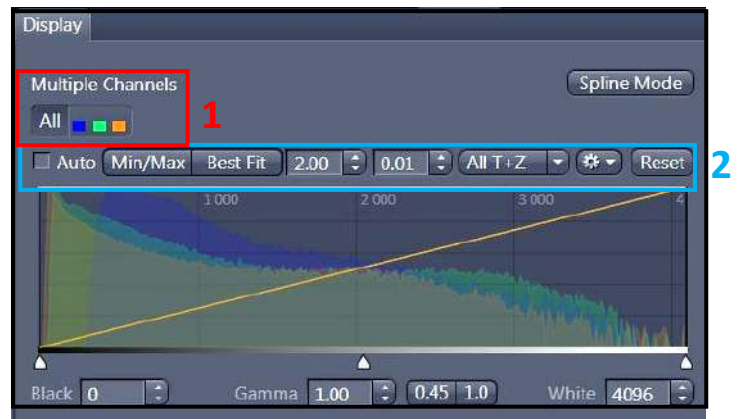
In the « *Dimensions* » Tab

1. Move in the Z axe or in the Time series by the displacement of the adapted.
2. Image size adjustment. With 100% 1 pixel in image acquisition corresponding to 1 pixel of the image obtained
3. Show / Hide the channel on the screen.
4. Saturation observation (Red is the saturated pixel).



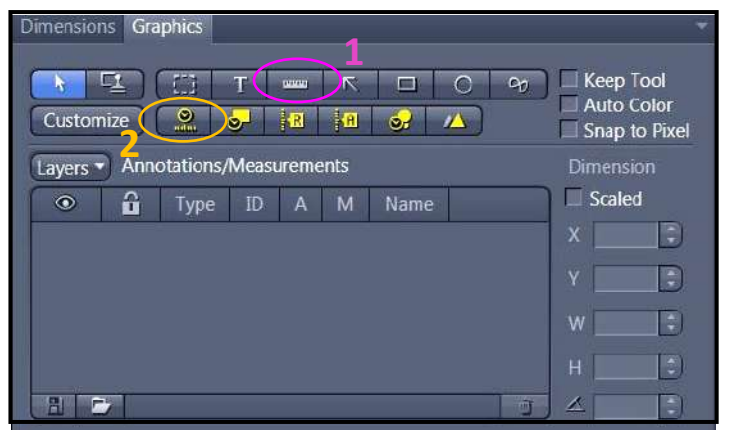
In the tab « *Display* »

1. Choose the channel or all to modify the image quality.
2. Contrast adjustment. Check on Reset to reinitialize it.



In Tab « *Graphics* »

1. Show the scale bar.
2. Show the time.



SIGNAL QUANTIFICATION

1. To quantify a fluorescence signal, you have to acquire them with the same acquisition parameters (same laser power, same gain, and Offset for PMTs).
2. To don't change the parameter you have to adjust your acquisition's settings with the sample whose fluorescence signal is the brightest. To limit the Over exposition effect.

IMAGE AND PARAMETER IMAGING

Settings Saving

1. select « Save As » in the « Parameter » logo. Name your settings and register its
2. to charge your parameter, open the menu « Experiment Manager » and choose your configuration.

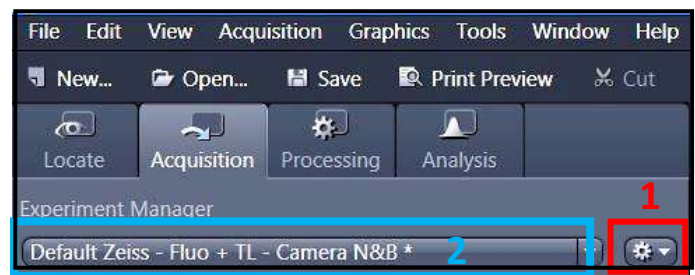
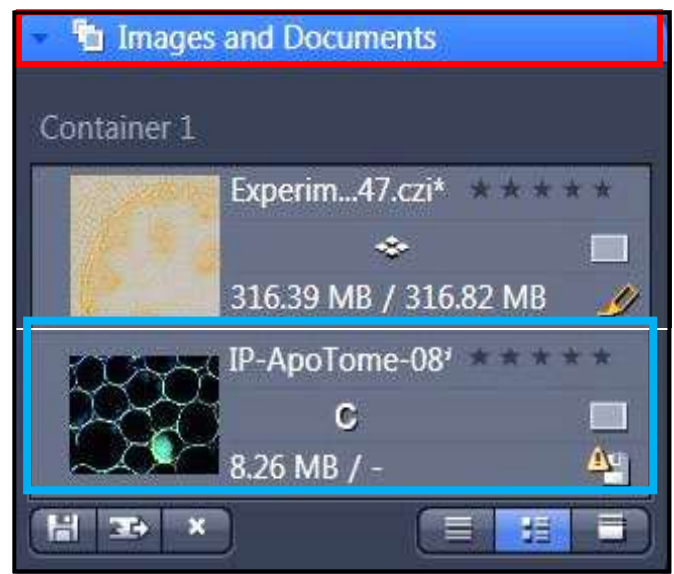


Image Saving

1. All images are visible on the right side called « Images and Documents ».
2. to save double-click on the images.
3. Click on the diskette logo, name your images, and save them in the folder « Users ».

Ps: create a folder with the date and your name.



STOP THE SYSTEM

1. Press « Components » button on ON
2. Press the button on the ON to activate the LED used to the ocular observation.
3. Switch on the PC and login into the session « USER ».
4. Start the software by clicking on the ZEN blue icon on the window desktop.
5. Click on the ZEN SYSTEM button to access the acquisition menu.



Module FLIM

I Start the System

1. Start the FRET / FLIM Computer

2. Start the software by clicking on the icon « Symphotime ».

3. On the LSM980 computer, open « ZenbluePicoQuantapp ».

4. Search the sample by the confocal mode to make your focus.

5. Select the « FLIM » detector in « Imaging Setup ».

6 Select the dichroic filter in the "Invisible light" section.

7. Select the « 485 » laser, it's the PicoQuant laser. Set to 100%.

8. Select one of the two lasers in the "Laser settings" tab of the "FLIM" window and choose the laser power.



The screenshot shows the FLIM software interface with several panels and settings:

- Top Panel:** A graph showing a spectral line at approximately 485 nm. The x-axis is labeled with 400, 500, 600, and 700. A dropdown menu is set to "Line".
- Channel Selection Table:**

Use	Dye	Color	Name	Range
<input type="checkbox"/>			Ch1	
<input type="checkbox"/>			ChS1	
<input type="checkbox"/>			Ch2	
<input type="checkbox"/>			ChA	
<input checked="" type="checkbox"/>			FLIM	5
<input type="checkbox"/>			T-PMT	
- Light Path Diagram:** Shows "Plate" and "Visible Light" components. A blue box highlights the "MBS T80/R20" dichroic filter and "Invisible Light" source, labeled with a green 6.
- Invisible Light Panel:** Shows "Lasers" with checkboxes for 405 and 485. The 485 nm laser is selected and highlighted with a green 7. Below it, a slider is set to 100.0%.
- Laser Settings Panel:** A blue header "Laser Settings" is followed by a table of laser parameters:

	Intensity	Power
<input checked="" type="checkbox"/> 442nm	1.0 %	70.0 %
<input type="checkbox"/> 481nm	1.0 %	65.0 %

The 442nm row is highlighted with a pink 8. Below this, an "All Lasers" slider is set to 100.0%.

II Application Settings

1. the frame size has to be **512 or 1024** not upper.
2. In the « Acquisition» tab, select « unidirectional ».
3. Select « Stop Manually » in « Measurement Settings ».
4. In the « PicoQuant » tab, set the « Repetition Rate » to 40MHz in « Laser Settings ».

This is the excitation frequency, it depends on the wavelength of the fluorescent labeling.

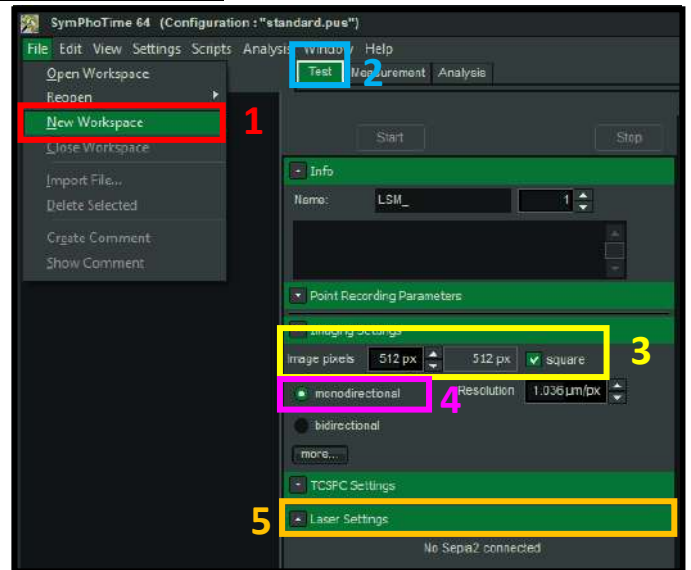
You need the excitation frequency allows seeing the fluorescence decay.

The screenshot shows the application settings interface with the following sections and settings:

- Image Size:** 499.1 μm \times 499.1 μm **Pixel Size:** 0.33 μm
- Frame Size:** 1515 px \times 1515 px (highlighted with a red box and callout 1). A "Presets" dropdown is also visible.
- Sampling:** 1.0 x Buttons: SR, Confocal
- Frame Time:** 44.68 s **Pixel Time:** 2.77 μs
- Scan Speed:** Slider and input field (5) Button: Max
- Direction:** 2 (highlighted with a blue box and callout 2). Options: Unidirectional (selected), Bidirectional.
- Averaging:** None, 2x, 4x, 8x, 16x
- Bits per Pixel:** 8, 16
- Application:** # FLIM [idle]
- Files:** Group: <Group>, File Name: <File>, Comment: <Comment>
- Measurement Settings:** 3 (highlighted with a green box and callout 3).
 - Stop Manually
 - Stop after Elapsed Time: 10.0 s
 - Photons in Brightest Pixel: 1000
 - Stop after Frames: 10
- Laser Settings:** 4 (highlighted with a pink box and callout 4).
 - Intensity:** 442nm (1.0%), 481nm (1.0%), All Lasers (100.0%)
 - Power:** 442nm (70.0%), 481nm (65.0%)
 - Repetition Rate:** 40.0 MHz
 - Laser Pattern:** Standard

In the « Symphotime » software during « Measurement »

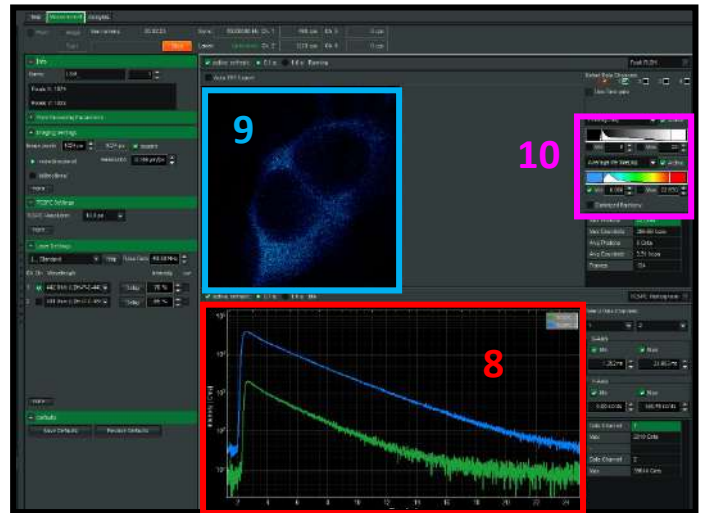
1. On the FLIM computer, in the "Symphotime" software, select «File» and create a new workspace.
2. Select the « Test » tab.
3. In « Imaging Settings », define the same pixel number as the frame size (see previously)
4. Choose the « Monodirectional » mode.
5. Select the same laser as the one chosen in Zen software.



6. In Zen software and start « test » in « Zeiss PicoQuant Application ». A set of parameters will have to be modified in order to obtain an optimal FLIM acquisition. This depends on the tagging, the medium, and the conditions.

7. in Zen, launch "Measurement" in the "Zeiss PicoQuant Application" tab

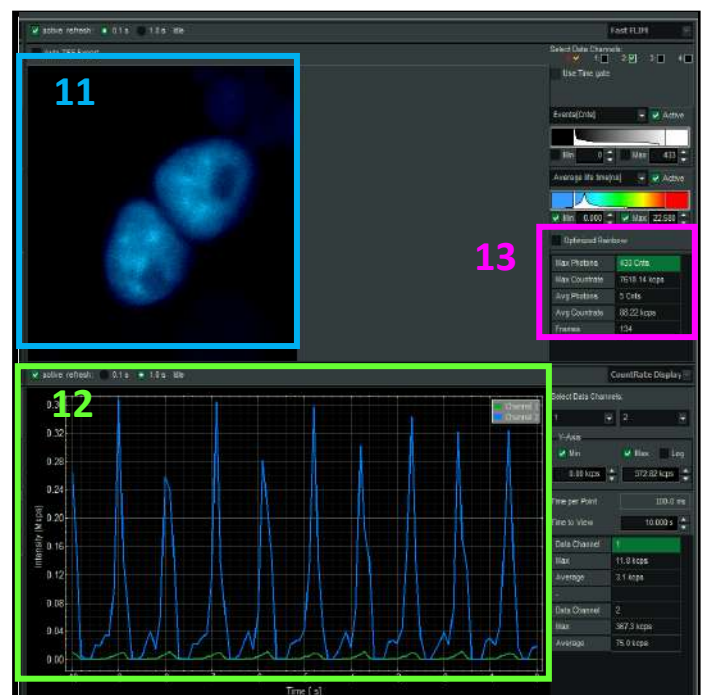
8. Fluorescence decay curve of detectors 1 and 2
9. Fluorescence lifetime Topography of detector 1
10. Fluorescence lifetime Histogram for each detector. And photons amount histogram.



11. Fluorescence lifetime Topography of detector 2

12. The maximum number of photons detected in the acquisition of a pixel. It will be the benchmark for stopping the experiment. **Count 1000 minimum to have a good signal for the analysis.**

13. In the Zen software, click on 'Stop' in the 'PicoQuant' tab to stop the acquisition.



Analysis of the FLIM acquisition

1. Go to « Analysis ».

2. In the « Imaging » tab, select « FLIM » mode and click on « Start ».

3. Select the acquisition folder.

In the new window, you will find a set of parameters and information about your FLIM Acquisition:

4. Fluorescence lifetime Histogram.

5. Fluorescence lifetime Topography.

6. Fluorescence decay curve (fluorescence intensity by time).

All of this information has been matched against the data set recovered during the acquisition. For further analysis please refer to the PicoQuant manual, and articles or contact the platform engineers directly.

